

Effect of modified atmosphere packaging, storage period, and storage temperature on the residual nitrate of sliced-pastırma, dry meat product, produced from fresh meat and frozen/thawed meat [☆]

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Abstract

The amount of nitrite in sliced-pastırma made, from fresh or frozen (which was stored at $-18\text{ }^{\circ}\text{C}$ for 240 days and then thawed at $10\text{ }^{\circ}\text{C}$ for 24 h) *M. Longissimus dorsi* muscle was determined. Sliced-pastırma samples were stored in modified atmosphere packages ($50\% \text{ N}_2 + 50\% \text{ CO}_2$) at 4 and $10\text{ }^{\circ}\text{C}$ for 150 days, and the amount of residual nitrite was measured after 0, 30, 60, 90, and 150 days of storage. The residual nitrite of pastırma samples made with frozen/thawed meat was higher than that of the pastırma made from fresh meat at both 0 day and at the end of the storage (150 days). The storage temperature ($p < 0.01$), storage period ($p < 0.01$) and the storage period \times the storage temperature interaction ($p < 0.01$) had significant effects on the amount of the residual nitrite. © 2004 Elsevier Ltd. All rights reserved.

Keywords: Pastırma; Residual nitrite; Modified atmosphere; Storage; Cured meat product

1. Introduction

Pastırma, a traditional Turkish meat product, is categorized (Gökalp, Kaya, & Zorba, 1999; Leistner, 1988) as an intermediate-moisture food (IMF). It is produced from whole muscle, obtained from certain parts of beef and water buffalo carcasses and, from one carcass, 16–20 different types of pastırma can be produced. In the pastırma process, muscles are cured, dried, pressed and

coated with a cement of garlic, red pepper, paprika, flour ground from seed of *Trigonella foenum graecum* and water. They are then dried again (maximum water content: 40%). Cement (paste seasoning) has a protective effect against mould growth and oxygen penetration, on and through the surface of the pastırma. High nutritional values and typical aromas from the cement have led to increase of pastırma consumption (Kaya, Aksu, & Gökalp, 1996; Tekinsen & Dogruer, 2000). Pastırma is the most popular dry-cured meat product in Turkey. A lot of dry-cured meat product is produced in the world, e.g., pastırma, bacon, Bündnerfleisch and ham, with or without a heat process. Cured-meat products differ greatly in composition and intended eating quality, but the types of bacteria growing on and in them are similar because the main factors controlling their growth are the same over a wide range of products

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(Roberts & Dainty, 1996; Topkim, 1980). Also, in cured meat products, curing may be combined with other processes, including drying, heating, smoking, and fermentation, but production of pastirma does not include heating or smoking processes.

The curing mixture used in pastirma production has a very important effect on pastirma, quality characteristics, so salt as well as nitrite and/or nitrate are used in the process of manufacturing pastirma. Also, another factor that influences quality is the penetration of curing compounds into the muscle. Numerous studies have shown that frozen/thawed meats can be penetrated more rapidly, since the salt penetrates faster than with fresh meats (Banon, Cayuela, Granados, & Garrido, 1999; Kemp, Langlois, & Johnson, 1982; Motilva, Toldra, Nadal, & Flores, 1994). In Turkey, fresh meat is usually used for pastirma production (Aksu & Kaya, 2001a, 2001b; Aksu, Aktas, & Kaya, 2002; Aksu & Kaya, 2005). However, Aksu, Kaya, and Ockerman (2005) reported that frozen/thawed meat could be used as a raw material. These researchers also determined that the quality characteristics of pastirma made from frozen/thawed meat are similar to those of pastirma made from fresh meat. Pastirma has to be sliced before being consumed without cooking. Preserving the quality characteristics of sliced pastirma can be a challenge, so sliced pastirma needs to be packaged in a modified atmosphere and stored at low temperature in order to prevent spoilage and to protect the sensory and microbiological quality.

The present study was conducted to determine the effect of modified atmosphere packaging, storage period, storage temperature and the use of fresh or frozen/thawed meat on the amount of residual nitrite remaining in sliced pastirma.

2. Materials and methods

2.1. Production of pastirma

M. Longissimus dorsi muscle from beef carcasses was used as the raw material in this study. The muscles were obtained from Eastern Anatolian Red beef carcasses (3-year-old cows) from a local slaughterhouse (Et ve Balik Kurumu Kombinasi, Erzurum, Turkey). For treatments (fresh and frozen/thawed), eight carcasses were used and one carcass was used for each replication. After removing fat and connective tissue from surfaces of the *Longissimus dorsi* muscle, the muscle was cut across the centre of the muscle into two pieces. Then half of the muscles were divided into two groups. One group was immediately used for pastirma production; the other group was frozen at -20°C and stored at -18°C for 240 days. The frozen muscles were then thawed at 10°C for 24 h and the muscles were used for pastirma pro-

Table 1
The stages of pastirma production (Aksu, 1999; Aksu & Kaya, 2002a)

| Production stage | Time (days and hours) | Temperature ($^{\circ}\text{C}$) | Relative humidity (%) |
|--|-----------------------|------------------------------------|--------------------------|
| Dry curing ^a | 2 days | 6 ± 0.5 | 80 ± 1 to 90 ± 1 |
| First drying | 4 days | 15 ± 0.5 | 80 ± 1 to 85 ± 1 |
| First pressing ^b | 17 h | 7 ± 0.5 | |
| Second drying | 3 days | 20 ± 0.5 | 70 ± 1 |
| Second pressing ^b | 7 h | 25 ± 0.5 | |
| Paste seasoning (cementing) ^c | 4 days | 7 ± 0.5 | |
| Third drying | 2 days | 15 ± 0.5 | 70 ± 1 |
| | 2 days | 18 ± 0.5 | 65 ± 1 |
| | 6–8 days | 20 ± 0.5 | 60 ± 1 |

^a For each 1 kg of meat, 50 g of curing mixture were used and the composition can be seen in Table 2.

^b For each 1 kg of meat, a 25 g weight was used.

^c 3–4 mm thickness of paste was used on the surface and the composition can be seen in Table 2.

Table 2

The composition of curing (Aksu, 1999; Aksu & Kaya, 2002a) and paste seasoning (cement) (Cankaya, 1997; Aksu, 1999) used in pastirma production

| Curing mixture composition ^a | | Cement mixture composition ^c | |
|---|---------------------|--|---------|
| NaCl | 47.25 g | Flour (<i>Trigonella foenum graecum</i> seed) | 500 g |
| KNO ₃ | 0.75 g | Smashed fresh garlic | 350 g |
| Glucose | 1.0 g | Paprika | 75 g |
| Sucrose | 1.0 g | Red pepper | 75 g |
| Starter Culture ^b | 25 g/100 kg of meat | Water | 1200 ml |

^a For each 1 kg of meat, 50 g of curing mixture were used.

^b A commercial preparation of *Staphylococcus carnosus* + *Lactobacillus pentosus* (Bactoferm™ C-P-77 S) was used as the starter culture (Chr Hansen's, Pohlheim, Germany), and starter culture was introduced into muscle with the curing mixture.

^c After the second drying step of pastirma production, the dried meat samples were pasted with the seasoning mix and the surface of the meat was covered (3–4 mm thickness) with the seasoning mixture.

duction. The production stages of pastirma are presented in Table 1. The curing mixture and composition of paste seasoning (cementing) used in pastirma production are shown in Table 2.

2.2. Packaging of sliced pastirma

Pastirma, produced from fresh or frozen/thawed meat, was sliced 1–2 mm in thickness using a slicing machine. The sliced pastirma samples were then packaged in 90 ± 5 g portions in a moderately gas impermeable OPAEVOH/PE (water vapour transmission rate $15 \text{ g/m}^2/24 \text{ h}/38^{\circ}\text{C}$, 90% RH, 1 atm; O₂ transmission rate $5 \text{ cm}^3/\text{m}^2/24 \text{ h}/23^{\circ}\text{C}$, 50% RH, 1 atm; N₂ transmission rate $1 \text{ cm}^3/\text{m}^2/24 \text{ h}/23^{\circ}\text{C}$, 50% RH, 1 atm; CO₂ trans-

mission rate $23 \text{ cm}^3/\text{m}^2/24 \text{ h}/23 \text{ }^\circ\text{C}$, 50% RH, 1 atm) bag. Packages were evacuated, filled with a modified atmosphere containing 50% N_2 and 50% CO_2 (the volume ratio of gas to sliced pastirma product was 1:2) and automatically heat-sealed by a Multivac packaging unit (Multivac A 300/16, Sepp Haggenmüller, D 87787 Wolfertschwenden, Germany).

2.3. Residual nitrite analysis

Residual nitrite was determined at 0, 30, 60, 90, 120, and 150 days of storage at 4 or 10 $^\circ\text{C}$. Residual nitrite was spectrophotometrically determined according to the methods of Anonymous (1981) and Tauchmann (1987). Ten-grammes of minced pastirma sample were homogenized using an Ultra-Turrax moderator (Nach Prof. P. Willem, Janke and Kunkel KG, IKA Werk, Staufen I. Breisgau), in 10 ml of saturated borax solution ($25 \text{ g Na}_2\text{B}_4\text{O}_7 \cdot 10\text{H}_2\text{O}$ for 500 ml deionized water) and 100 ml of boiling deionized water; then this mixture was placed in a bag to boil in a water bath for 15 min, and cooled with tap water for 3 min. The content of this mixture was transferred into a 200 ml volumetric flask 2 ml of Carrez-I ($26.5 \text{ g K}_4(\text{Fe}(\text{CN})_6 \cdot 3\text{H}_2\text{O})$) into 250 ml deionized water) were added and also 2 ml of Carrez-II [$55 \text{ g Zn}(\text{CH}_3\text{COO})_2 \cdot 2\text{H}_2\text{O} + 7.5 \text{ ml}$ glacial acetic acid (99.5 %) into deionized 250 ml water] solution were also added. The total 200 ml volume was adjusted by adding deionized water. This mixture was allowed to settle at room temperature (20 $^\circ\text{C}$) for 30 min and then filtered through Whatman No: 42 filter paper. Ten ml of the filtrate were combined with, 5 ml of sulfanilamide solution [$1.5 \text{ g NH}_2\text{C}_6\text{H}_4\text{SO}_2\text{NH}_2$ were added to 62.5 ml HCl (37%) and made up to 250 ml with deionized water] and 5 ml of *N*(1-naphthyl)-ethylenediaminedihydrochloride solution [$0.25 \text{ g C}_{12}\text{H}_{14}\text{N}_2 \cdot 2\text{HCl}$ for 250 ml volume with deionized water], and this mixture was allowed to settle at room temperature (20 $^\circ\text{C}$) for 30 min. At the end of 30 min, absorbances of the solutions were determined at 540 nm (utilizing a blank without the sample solution) with a spectrophotometer (Shimadzu, UV-160A UV-Visible Recording Spectrophotometer). A standard curve was prepared using pure sodium nitrite (NaNO_2). Results were expressed as ppm of NaNO_2 .

Changes (%) in the amount of residual nitrite were calculated by differences at various storage days and the results are shown in Table 3 as ppm of residual nitrite and as percentage (%) change when compared to day 0.

2.4. Statistical analysis

Experimental data were statistically processed, using the SPSS version, with 10.01 software (SPSS, 1996). Data were analyzed as a completely randomized design procedure, and comparisons of treatment means were

Table 3

The amount of residual nitrite (ppm) \pm standard deviation of sliced and packaged (50% N_2 + 50% CO_2) pastirma samples during storage

| Storage period (day) | Frozen/thawed | | Fresh | |
|----------------------|------------------------|--------------------------------|------------------------|--------------------------------|
| | Residual nitrite (ppm) | Changes compared to 0 days (%) | Residual nitrite (ppm) | Changes compared to 0 days (%) |
| 0 | 22.42 \pm 0.57 a | 0.00 | 11.60 \pm 0.10 a | 0.00 |
| 30 | 19.82 \pm 1.52 b | 11.60 | 11.49 \pm 1.23 a | 0.95 |
| 60 | 15.04 \pm 1.58 c | 32.92 | 9.20 \pm 0.37 b | 20.69 |
| 90 | 8.15 \pm 0.92 d | 63.65 | 3.45 \pm 0.14 c | 70.26 |
| 120 | 7.59 \pm 0.87 d | 66.15 | 3.00 \pm 0.15 cd | 74.14 |
| 150 | 5.27 \pm 1.62 e | 76.49 | 2.57 \pm 0.18 d | 77.84 |
| P | --^{A} | | --^{A} | |

P: Probability.

a–e: Means with different letters in the same column are significantly different at ($P < 0.05$).

^A $P < 0.01$ (fresh vs frozen/thawed).

Table 4

The amount of residual nitrite (ppm) \pm standard deviation for pastirma stored at 4 and 10 $^\circ\text{C}$

| Storage temperature | Frozen/thawed | Fresh |
|---------------------|------------------------|------------------------|
| 4 $^\circ\text{C}$ | 13.55 \pm 6.29 a | 6.63 \pm 3.98 b |
| 10 $^\circ\text{C}$ | 12.55 \pm 7.35 b | 7.14 \pm 4.34 a |
| P | --^{A} | --^{A} |

P: Probability.

a–b: Means with different letters in the same column are significantly different at ($P < 0.05$).

^A $P < 0.01$ (fresh vs frozen/thawed).

based on Duncan's multiple range test. Mean values and standard deviation (\pm) are shown in Tables 3 and 4.

3. Results and discussion

Table 3 shows the mean values and standard deviations, for residual nitrite and changes for each storage period (30, 60, 90, 120, and 150 days) compared to 0 day for the pastirma samples. Pastirma produced from frozen/thawed meat had higher ($p < 0.01$) residual nitrite than that produced from fresh meat at 0 days of storage, although the processes, of pastirma (fresh and frozen/thawed) production used the same curing mixture and the same curing time (Table 3). This difference may be explained by the properties of the raw material used for pastirma production. It has been reported, in related research, that frozen/thawed meat allows more rapid penetration of curing matter (Banon et al., 1999; Kemp et al., 1982; Motilva et al., 1994). In sliced-pastirma, produced from frozen/thawed meat, the highest amount of residual nitrite (22.4 ± 0.57 ppm) was observed in the early days of storage (0 days). This was significantly

decreased (5.27 ± 1.62 ppm) during storage. A significant decrease at 150 days of storage, by 77.84% compared to day 0 (Table 3), was also observed in sliced-pastirma produced from fresh meat.

There was a decrease ($p < 0.01$) in the amount of residual nitrite of sliced pastirma (fresh and frozen/thawed) during storage. The lowest residual nitrite values was found at 150 days in sliced pastirma produced from frozen/thawed meat, while lower residual nitrite values were observed after 90–150 days of storage for sliced pastirma produced from fresh meat (Table 3). There was also a statistically significant ($p < 0.01$) decrease in nitrite values between 90 and 150 days of storage for pastirma produced from both fresh and frozen/thawed meat (Table 3).

The residual nitrite values of sliced and packaged pastirma in modified atmosphere samples stored at 4 and 10 °C are shown in Table 4. The storage temperature (4 and 10 °C) had an effect ($p < 0.01$) on the amount of residual nitrite. Higher residual nitrite value was found in the pastirma produced from frozen/thawed meat and the samples stored at 4 °C had higher residual nitrite values than those stored at 10 °C (Table 4).

An interaction ($p < 0.01$) of the storage period \times the storage temperature was found for residual nitrite of both pastirma groups (fresh and frozen/thawed), as shown in Fig. 1. This figure illustrates that the fastest decrease in residual nitrite, among both pastirma groups, occurred between 30 and 90 days of storage.

Nitrite, if used improperly, can be dangerous from a human health standpoint; hence, the level of usage is limited, and in some countries, the use of nitrite has been banned. It is also advisable to use a mixture of salt with nitrite to reduce accidental formulation mistakes. Nitrite

and/or nitrate need to be added to the curing mixture for cured meat products to enhance colour, and flavour, reduce bacterial growth and reduce oxidation properties of the final product (Gökalp et al., 1999; Kaya, 1995; Kaya et al., 1996; Wirth, 1986; Prändl, Fischer, Schmidhofer, & Sinell, 1988). According to the Turkish Food Additive Codex (Anonymous, 1997), the maximum amount of residual nitrite for cured meat products is 50 mg/kg. In the present study, the determined residual nitrite values in all the pastirma samples, produced by the current procedure from fresh and frozen/thawed meat were below this level (< 50 mg/kg, NaNO_2). There has been some research on the amount of residual nitrite of pastirma produced in Turkey. For instance, El-Khateib, Schmidt, and Leistner (1987) reported that the amount of residual nitrite in Turkish pastirma ranged from 2 to 58 mg/kg (average: 12 mg/kg). Soyutemiz and Özenir (1996) found that the amount of residual nitrite in pastirma was 16.0 mg/kg of product found in the market place in Bursa. Cankaya (1997) reported that the amount of residual nitrite in pastirma was 0.43–6.37 mg/kg. Yagli (Gür) and Ertas (1998) made similar observations (13.4–25.8 mg/kg) during investigations of products also containing sodium ascorbate in the production of pastirma. Tyrpenou, Gouta, Tsigouri, and Vlasiotis (2000) reported that the amount of residual nitrite in Greek pastirma ranged from 0.85 to 190 mg/kg as sodium nitrite. Aksu and Kaya (2001a) reported that the amount of residual nitrite of pastirma marketed in Erzurum was 0.93–11.6 mg/kg. Aksu and Kaya (2002a, 2002b) also reported that the amounts of residual nitrite in pastirma produced by different curing methods (dry- and brine-curing) and pastirma produced using potassium nitrate and a starter culture were 44.1–

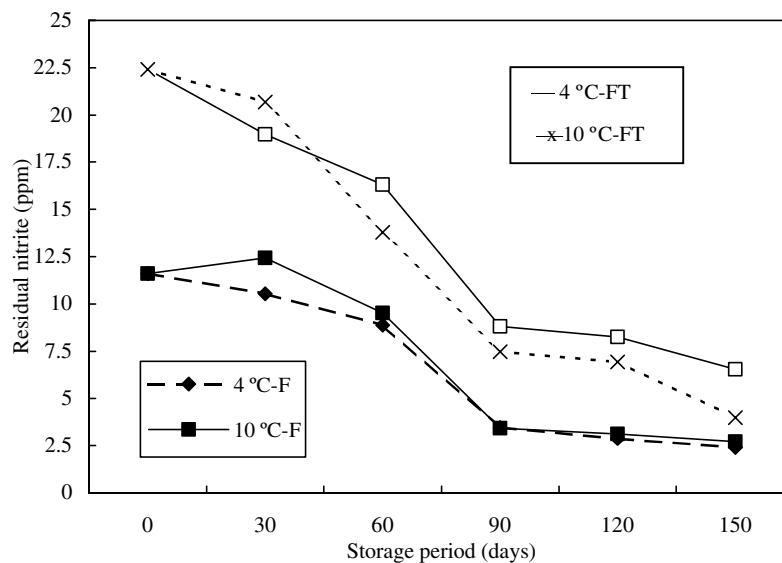


Fig. 1. Effect of fresh and frozen/thawed raw material, storage period and storage temperature on the residual nitrite content of pastirma samples (Raw material for pastirma: F, Fresh meat and FT, Frozen/Thawed meat).

47.9 and 64.3–75.6 mg/kg, respectively. Dogruer, Guner, Gurbuz, and Ucar (2003) reported that the residual nitrite of pastirma produced by using sodium and potassium nitrate was 22.1–143 mg/kg. Aksu and Kaya (2001b, 2002c) in another studies, determined that the use of commercial starter cultures in pastirma production, decreased the amount of residual nitrite [8.50–21.3 mg/kg (2001b); 22.7 mg/kg (2002c)]. Furthermore, various researchers have reported that use of starter cultures, in dry-cured raw meat products like pastirma, increased the quality properties of the final (end) product (Aksu & Kaya, 2001b, 2002a, 2002b; Aksu et al., 2002; Katsaras, Lautenschlager, & Bosklova, 1996). Some types (*Micrococcaceae*, such as *Staphylococcus carnosus*, *Staphylococcus xylosum*, etc) generally contain nitrate reductase, and proteolytic and lipolytic enzymes (Johansson, Berdague, Larsson, & Borch, 1994; Krockel, 1995), and these microorganisms can be used as starter cultures in cured meat products (Hammes & Hertel, 1998). Nitrate has to be converted to nitrite to improve the quality properties of meat products (Geisen, Lücke, & Kröckel, 1992; Lücke, 1985). This conversion is enzymatic and due to the activity of nitrate reductase which is an intracellular enzyme (Jessen, 1995). Also, Geisen et al. (1992) reported that nitrite is also reduced by *Micrococcus* and *Staphylococcus*; therefore, the amount of nitrite in the final products is lower than in products produced without *Micrococcus* and *Staphylococcus*. In addition, these microorganisms (starter cultures) grow easily in muscle tissue, because they are facultative anaerobes or have anaerobic properties. Katsaras et al. (1996) have reported that the *Micrococcus/Staphylococcus* count in the centre of muscle, at the end of curing, is increased (5.0–7.53 log CFU/g) due to the starter cultures (*S. carnosus*, *L. curvatus* and *M. Varians*) utilized in pastirma production. Also, Aksu (1999) and Aksu and Kaya (2002a) reported that the *Micrococcus/Staphylococcus* and lactic acid bacteria counts in samples with starter culture, at the end of curing, of the pastirma process, were higher than without starter culture. Also, Hammes and Knauf (1993) reported that lactic acid bacteria promoted activity of nitrate reductase. *L. plantarum* and *L. pentosus* have higher nitrate reductase activity than other lactic acid bacteria (Hammes & Knauf, 1993; Jessen, 1995; Krockel, 1995; Wolf & Hammes, 1988). This study showed that nitrate, both near the surface and in the centre of the muscle, was reduced by the addition of the starter cultures. In frozen/thawed meats, the penetration of the curing mixture has been shown to be more rapid than fresh meats, because pastirma produced from frozen/thawed meat was determined to have higher residual nitrite than that produced from fresh meat (Table 3). The structure of muscle tissue is damaged by freezing and thawing. Muscle proteins are also denatured by freezing and thawing (James & James, 2002), and so

the curing mixture (both salt and nitrate and starter cultures) penetrates faster in frozen/thawed muscle tissues than in intact muscles.

Gas dissolution, which causes changes during pastirma aging, depends on the storage temperature and properties of the gas. Gases such as carbon dioxide (CO₂) and nitrogen (N₂) helps to maintain the quality of some foods during storage. Many gases and gas combinations are used, depending on the kind and properties of the meat products. Carbon dioxide (CO₂) and/or nitrogen (N₂) combinations are usually used for processed meat products. Carbon dioxide has the most significant effect on the food shelf life because of the increased dissolution of CO₂ at low temperatures. Jo, Ann, Son, Lee, and Byun (2003) reported that the amount of residual nitrite of sausage samples packaged with CO₂ (100%) was lower than that with vacuum and aerobic packaging, and the redox potential changes to a reduced state in vacuum or CO₂ packaging and can increase the possibility of transforming the nitrite ion to nitric oxide, resulting in a reduction of residual nitrite in sausages.

4. Conclusion

The results from the present study indicate that there is no risk due to residual nitrite associated with using frozen/thawed meat for pastirma manufacturing. This research suggests that the curing time (in process) for frozen-thawed meat may be shortened, and/or less nitrate can be used in the curing and storage of this product. However, from the results, it was observed that additional experiments are needed to determine the most appropriate curing time and initial nitrate concentration, when frozen-thawed meat is used for pastirma manufacturing.

References

- Aksu, M. I. (1999). *Research on the possibility of starter culture use in pastirma production* (Pastirma uretiminde starter kultur kullanim imkanlari) Ph.D. Thesis. Atatürk Univ. Graduate Institute of Science, Erzurum, Turkey.
- Aksu, M. I., & Kaya, M. (2001a). Some microbiological, chemical and physical characteristics of pastirma marketed in Erzurum. *Turkish Journal of Veterinary & Animal Science*, 25(3), 319–326.
- Aksu, M. I., & Kaya, M. (2001b). The effect of starter culture use in pastirma production on the properties of end product. *Turkish Journal of Veterinary & Animal Science*, 25(6), 847–854.
- Aksu, M. I., & Kaya, M. (2002a). Production of pastirma with different curing methods and using starter culture. *Turkish Journal of Veterinary & Animal Science*, 26(4), 909–916.
- Aksu, M. I., & Kaya, M. (2002b). The possibilities for the use of commercial starter cultures in pastirma production. *Turkish Journal of Veterinary & Animal Science*, 26(4), 917–923.

- Aksu, M. I., & Kaya, M. (2002c). Effect of commercial starter cultures on the fatty acid composition of pastirma (Turkish dry meat product). *Journal of Food Science*, 67(6), 2342–2345.
- Aksu, M. I., Aktas, N., & Kaya, M. (2002). Effect of commercial starter cultures on the myofibrillar protein of pastirma. *Journal of Food Science*, 67(7), 2548–2551.
- Aksu, M. I., & Kaya, M. (2005). Effect of storage temperatures and time on shelf life of sliced and modified atmosphere packaged-Pastirma, a dry meat product, produced from beef meat. *Journal of the Science of Food and Agriculture*, in press.
- Aksu, M. I., Kaya, M., & Ockerman, H. W. (2005). Effect of modified atmosphere packaging on the shelf life of sliced-pastirma produced from frozen/thawed meat. *Journal of Muscle Foods*, in press.
- Anonymous. (1981). Bestimmung des Nitrit- und Nitratgehaltes in Fleisch und Fleischerzeugnissen, Amtliche Sammlung von Untersuchungsverfahren nach 35 LMBG, Germany.
- Anonymous. (1997). *Food Codex of Turkish* (Türk Gıda Kodeksi). Tarım ve Köyisleri Bakanlığı. T.C. Resmi Gazete. Sayı: 23172, s: 44. Basbakanlık, Ankara, Turkey.
- Banon, S., Cayuela, J. M., Granados, M. V., & Garrido, M. D. (1999). Pre-cure freezing affects proteolysis in dry-cured hams. *Meat Science*, 51, 11–16.
- Cankaya, H. (1997). *The effects of calcium chloride on some quality and technological properties of pastirma* (Kalsiyum klorürün pastirmanın bazı kalite ve teknolojik özelliklerine etkisi, Yüksek Lisans Tezi. Atatürk Üniv. Fen Bilimleri Enstitüsü. Erzurum). Atatürk Univ. Graduate Institute of Science, Erzurum, Turkey.
- Dogrue, Y., Guner, A., Gurbuz, U., & Ucar, G. (2003). The effect of sodium and potassium nitrate on the quality of Turkish Pastrami (pastirma). *Turkish Journal of Veterinary & Animal Science*, 27, 805–811.
- El-Khateib, T., Schmidt, U., & Leistner, L. (1987). Microbiological stability of Turkish pastirma. *Fleischwirtschaft*, 67(1), 101–105.
- Geisen, R., Lücke, F., & Kröckel, L. (1992). Starter and protective cultures for meat and meat products. *Fleischwirtschaft*, 72(6), 894–898.
- Gökalp, H. Y., Kaya, M., & Zorba, O. (1999). Technology of pastirma and some other dried products. Engineering of Meat Products Processing. (3rd Press). Atatürk Univ. Publ. No: 786 Faculty of Agric. No: 320. Erzurum, Turkey, pp. 309–339.
- Hammes, W. P., & Knauf, H. J. (1993). Starters in the processing of meat products. *Meat Science*, 36, 155–168.
- Hammes, W. P., & Hertel, C. (1998). New developments in meat starter cultures. *Meat Science*, 49(1), 125–138.
- James, S. J., & James, C. (2002). *Meat refrigeration*. Abington Hall, UK: Woodhead Publishing Limited.
- Jessen, B. (1995). Starter cultures for meat fermentation. In Compell-Platt & P. E. Cook (Eds.), *Fermented meats*. New York, USA: Blackie Academic and Professional.
- Jo, C., Ann, H. J., Son, J. H., Lee, J. W., & Byun, M. W. (2003). Packaging and irradiation effect on lipid oxidation, color, residual nitrite content, and nitrosamine formation in cooked pork sausage. *Food Control*, 14(1), 7–12.
- Johansson, G., Berdague, J. L., Larsson, N. T., & Borch, E. (1994). Lypolysis, proteolysis and formation of volatile components during ripening of fermented sausage with *Pedococcus pentosaceus* and *Staphylococcus xylosus* as starter cultures. *Meat Science*, 38, 203–218.
- Katsaras, K., Lautenschlager, R., & Bosklova, K. (1996). Verhalten von mikroflora und starterkulturen während der Pokelung, Trocknung, und Lagerung von Pasterma. *Fleischwirtschaft*, 76(3), 308–314.
- Kaya, M. (1995). Sucuk, Pastirma ve Kavurmanın Sağlık Açısından İrdelenmesi, *Standard* (Özel Sayı). s:65–68.
- Kaya, M., Aksu, M. I., & Gökalp, H. Y. (1996). Dry curing-raw meat products. In *Symposium of Meat and Meat Products '96*. Istanbul Univ. Vet. Fac. Istanbul, Turkey, pp. 26–34.
- Kemp, J. D., Langlois, B. E., & Johnson, A. E. (1982). Effect of pre-cure freezing and thawing on the microflora, fat characteristics and palatability of dry-cured ham. *Journal of Food Protection*, 45, 244–248.
- Krockel, L. (1995). *Bacterial fermentation of meats. Fermented meats* (pp. 69–102). New York: Blackie Academic and Professional.
- Leistner, L. (1988). *Hurdle technology in meat products and other foods*. In Stufe R. (Ed.), *Lebensmittelqualität, Wissenschaft und Technik*. Wissenschaftliche Arbeitstagung, 25 Jahre Institut für Forschung und Entwicklung der Maizena GmbH; March 2–4, 1988 Heilborn, Germany, pp. 58–63.
- Lücke, F. K. (1985). Mikrobiologische Vorgänge bei der Herstellung von Rohwurst und Roschinken. In: *Mikrobiologie und Qualität von Rohwurst und Roschinken*. Bundesanstalt für Fleischforschung, Kulmbach, pp. 85–102.
- Motilva, M. J., Toldra, F., Nadal, M. I., & Flores, J. (1994). Pre-freezing hams affects lipolysis during dry-curing. *Journal of Food Science*, 59(2), 303–305.
- Prändl, O., Fischer, A., Schmidhofer, T., & Sinell, H. J. (1988). *Fleisch-Technologie und Hygiene der Gewinnung und Verarbeitung*. Stuttgart, Germany: Verlag Eugen Ulmer.
- Roberts, T. A., & Dainty, R. H. (1996). Nitrite and nitrate as food additives: rationale and mode action. In M. Hill (Ed.) *Nitrates and nitrites in food and water* (pp. 113–124, Chapter 6). Abington Hall, Abington, Cambridge, UK: Woodhead Publishing Limited.
- Soyutemiz, G. E., & Özenir, A. (1996). Determination of residual nitrate and nitrite contents of dry fermented sausage, salami, sausage and pastirma consumed in Bursa (Bursa'da Tüketilen Sucuk, Salam, Sosis ve Pastirma'lardaki Kalinti Nitrat ve Nitrit Miktarlarının Saptanması). *Gıda*, 21(6), 471–476.
- SPSS (1996). *SPSS users guide*. Windows Release 10.01, SPSS Inc.
- Tauchmann, F. (1987). Methoden der chemischen Analytik von Fleisch und Fleischwaren. (p. 80). Bundesanstalt für Fleischforschung, Kulmbach, Germany.
- Tekinsen, O. C., & Dogruer, Y. (2000). *Pastirma. From Every Aspects*. Konya, Turkey: Selcuk Univ. Press.
- Topkim, R. B. (1980). Botulism from meat and poultry products – a historical perspective. *Food Technology*, 34, 229–257.
- Tyrpenou, A. E., Gouta, E. H., Tsigouri, A. D., & Vlasiotis, C. N. (2000). Nitrite and nitrate residual in Greek Pastirma. Deltion tes Ellenikes Kteniatrikes Etaireias. *Bulletin of the Hellenic Veterinary Medical Society*, 51, 302–307.
- Wirth, L. (1986). Zur Technologie bei rohen Pökelfleischerzeugnissen. *Fleischwirtschaft*, 66(4), 531.
- Wolf, G., & Hammes, W. P. (1988). Effect of hematin on the activities of nitrite reductase and catalase in Lactobacilli. *Arch Microbiology*, 149, 220–224.
- Yaglı (Gür), H., & Ertas, A. H. (1998). Effect of sodium ascorbate on some quality characteristics of Turkish pastirma. *Turkish Journal of Agriculture & Forestry*, 22, 515–520.